

## A Review on Herbal Plants with Hepatoprotective Effects

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### Abstract

The development of liver illnesses has emerged as a serious concern in world health. These diseases may be caused by several harmful compounds, such as chemotherapeutic drugs, thioacetamide, carbon tetrachloride, certain antibiotics, heavy alcohol use, and harmful bacteria. Therefore, excellent health and well-being depend on maintaining a functioning liver. Synthetic drug drawbacks have surpassed their benefits, despite pharmacological advancements. Contemporary medical approaches to liver disease treatment are not only inefficient, but also linked to harmful side effects and very high prices, making them unaffordable for underdeveloped nations. Therefore, there seems to have been a global upsurge in interest in investigating medicinal plants as potential alternative treatment agents for illnesses. These plants are readily accessible, inexpensive, and do not need labor-intensive pharmaceutical manufacturing methods. The focus has so been on traditional herbs that are very effective, have minimal toxicity, and are cost-efficient. This research presents the results of a literature search that used many databases, including PubMed, ISI Web of Knowledge, and Google Scholar. We reviewed all available material on medicinal plants from throughout the globe that have the potential to protect the liver. Additionally, we covered phytochemical substances that have hepatoprotective properties, and we concluded by highlighting upcoming research in the topic.

**Key words:** Bioactive compounds, hepatotoxicity, liver diseases, medicinal plant, pharmacology.

### Introduction

The liver, with its many roles in the body, is the most important and vital organ. In addition to excreting waste metabolites, it plays a role in the metabolism of nutrients including lipids, proteins, and carbs. The liver has a role in the breakdown and removal of toxins, including medicines and other foreign chemical compounds, because it is the first organ in the body to receive toxins from the intestines. The liver's precise roles in the body are as follows.

There are a lot of chemicals that may be stored in it, including glycogen, minerals, vitamins, and iron. When energy needs arise and blood sugar drops too low, the liver converts glycogen stores into glucose, which the body may use. Toxin by-products, pathogenic organisms,

medications, alcohol, chemicals, heavy metals, and more are all removed from the bloodstream by this process. In addition, chemicals, medications, viruses, bacteria, parasites, fungus, herbicides and pesticides, lipids, dietary additives, alcohol, and dead cells are among the waste products and pollutants of blood that the liver eliminates. Furthermore, the liver is sometimes referred to as the body's biochemical unit due to the fact that it performs all duties via various organs including the lungs, skin, and mouth. Prior to their distribution to various areas of the body, it metabolises chemicals in the circulation. Digesting and emulsifying lipids, oils, and other substances (including vitamins A, D, E, and K) requires the production of bile,

which the liver produces as part of its digestive function [4]. Blood proteins, enzymes, hormones, immunological factors, and clotting factors are among the proteins that it may produce. At last, during times of low blood sugar, the liver may manufacture cholesterol, a transporter for the energy-supplying lipids required for adenosine triphosphate production in the body.

#### HEPATOTOXICITY AND LIVER DISEASES

The incidence of liver illnesses in developing nations ranks high among the world's most pressing health concerns. Different types of liver illness include noninflammatory hepatitis, inflammatory acute or chronic hepatitis, and degenerative cirrhosis or fibrosis. They are usually caused by heavy metals, poisons, starvation, and the use of OTC drugs without a doctor's prescription. Alcoholic liver disease, hepatitis, jaundice, and liver fibrosis are the end outcomes of the aforementioned causes' destruction and impairment of hepatocytes. One sign of liver damage or illness is an increase in blood cholesterol levels. Cardiovascular disorders are more likely to occur in individuals with high levels of low-density lipoprotein cholesterol (LDL-C) and triacylglycerols (TAGs).[5]

Further factors that have been extensively studied and found to cause damage to hepatic cells include hepatic cell overconsumption, toxic substances like thioacetamide (TAA), drug abuse (e.g., paracetamol), chemotherapeutic agents like carbon tetrachloride (CC14), aflatoxin, microbes, and viral infections (e.g., hepatitis A, B, C, and D). From 6 to 8, A chain reaction occurs and potentially initiates lipid peroxidation when the endoplasmic reticulum and mitochondrial cytochrome P-450 metabolise CC14, leading to the generation of reactive oxygen species (ROS,  $CC13O^-$ ).

PCM is often used as an analgesic or an antipyretic to reduce or eliminate fever. Overdosing on this medication causes harm to the liver cells, which in turn causes illness or injury to the liver.[9] Moreover, considerable excessive hepatic damage may result from the death of the majority of liver cells (necrosis), which is marked by nuclear pyknosis and eosinophilic cytoplasm, when PCM is administered in excess. An oxidative byproduct,

N-acetyl-P-benzoquinoneimine, is formed during PCM metabolism in the liver. This compound makes a covalent connection with the sulfhydryl groups of proteins, namely with cytochrome P-450 enzymes. Hepatocyte necrosis results from this process's ultimate cause: the peroxidative breakdown of glutathione (GSH) lipids.

One other chemical that causes membrane damage is trimethylammonium (TAA), which blocks the free passage of RNA between the nucleus and cytoplasm. This damage to the liver is caused by the TAA metabolite.[9] In addition to lowering the frequency of oxygen consumption, TAA may decrease the number of hepatocytes. Furthermore, it decreases both the amount of bile and the concentration of bile salts and deoxycholic acid within it. An rise in blood levels of toxins is a symptom of hepatotoxin-associated liver damage, which causes abnormal bile excretion.[10] The health of humans depends on this liver's ability to function normally at all times. From 11 to 13, Despite its remarkable regenerating capabilities, the liver is constantly exposed to harmful environmental contaminants such xenobiotics and chemotherapeutic drugs, which may inhibit its natural defensive mechanisms and cause liver dysfunction and damage.[14]

Conversely, most hepatotoxic substances cause damage to hepatocytes, which in turn hinders kidney function, often via oxidative processes such lipid peroxidation. The body's antioxidant mechanisms fall short when the liver is injured. Radon oscillations (ROS) may be produced by several external factors, including X-rays, contaminants, UV radiation, or metabolic reactions inside the mitochondria.[15] The rate of ROS generation and clearance by various endogenous antioxidants, including enzymatic and nonenzymatic mechanisms, is the only factor that determines the intracellular concentration of ROS.[15]

According to many studies, free radicals cause oxidative stress, which in turn causes hepatocyte degeneration, swelling, necrosis, and apoptosis. Lipid peroxidation and covalent binding are the typical pathways by which free radicals cause liver damage or damage, leading to subsequent tissue destruction. The lipids, proteins, and nucleic acid in cell membranes are destroyed by reactive oxygen species (ROS),

which have been associated with various age-related problems such as atherosclerosis, diabetes, kidney and lung damage, liver disorders, cancer, inflammatory diseases, and cardiovascular diseases. On pages 16 and 17, Cell membranes are vulnerable to lipid peroxidation, which compromises their structural integrity and functioning. This, in turn, reduces the cell's ability to sustain consistent ion gradients and transport.[18] Conversely, chemical exposure and excessive drug usage may also harm the liver.[14] The effects of many medicines on the liver have been documented, as shown in Table 1 and Figure 1.

**FREE RADICALS AND LIPID PEROXIDATION**

To prevent free radical-induced lipid peroxidation, the free radical scavenging process is crucial. The metabolic pathway that begins with ethanol exposure enhances lipid peroxidation, which in turn causes hepatitis and, ultimately, cirrhosis [19].

In recent decades, hepatoprotective medicines derived from less toxic plant compounds have been used. Therefore, there has been significant study in this sector focused on continuously exploring plant diversities for new hepatoprotective potential. In [20], It is important to maintain a balance between reactive oxygen species (ROS) and antioxidant enzymes like glutathione peroxidase (GSH-Px), superoxide dismutase (SOD), and catalase

(CAT) in order to prevent damage caused by oxidative stress, as earlier research has shown that an excess of ROS intensifies oxidative stress, leading to health problems like diabetes, kidney and liver injury, cancer, and heart disease. Natural lipid peroxidation defenders, the enzymatic antioxidant defence systems [22] include Cu-Zn, Mn-SOD, CAT, and GSH reductase; they work by directly or sequentially removing ROS, therefore halting or decreasing this process.[23] Glutathione (GSH) is a crucial cytosolic antioxidant that plays a role in the detoxification and excretion of xenobiotics; keeping its level high is vital for avoiding lipid peroxidation. CCl4 is one of the xenobiotics that may cause acute damage to liver cells by generating free radicals, namely trichloromethyl radicals [24].[25] Typically, the liver has an increasingly protective mechanism for compounds that enhance the activity of glutathione S-transferase, an enzyme that may transform harmful molecules into innocuous ones. Because they have less of an impact on the body's processes, natural goods, such as medicinal plants and their constituents, have the potential to cure and prevent a wide range of disorders. references 26 and 27 Research has shown that some herbal extracts may have a protective effect on an overloaded liver. 8,13, and 28

**Table 1: Example of some drugs with hepatotoxicity effects**

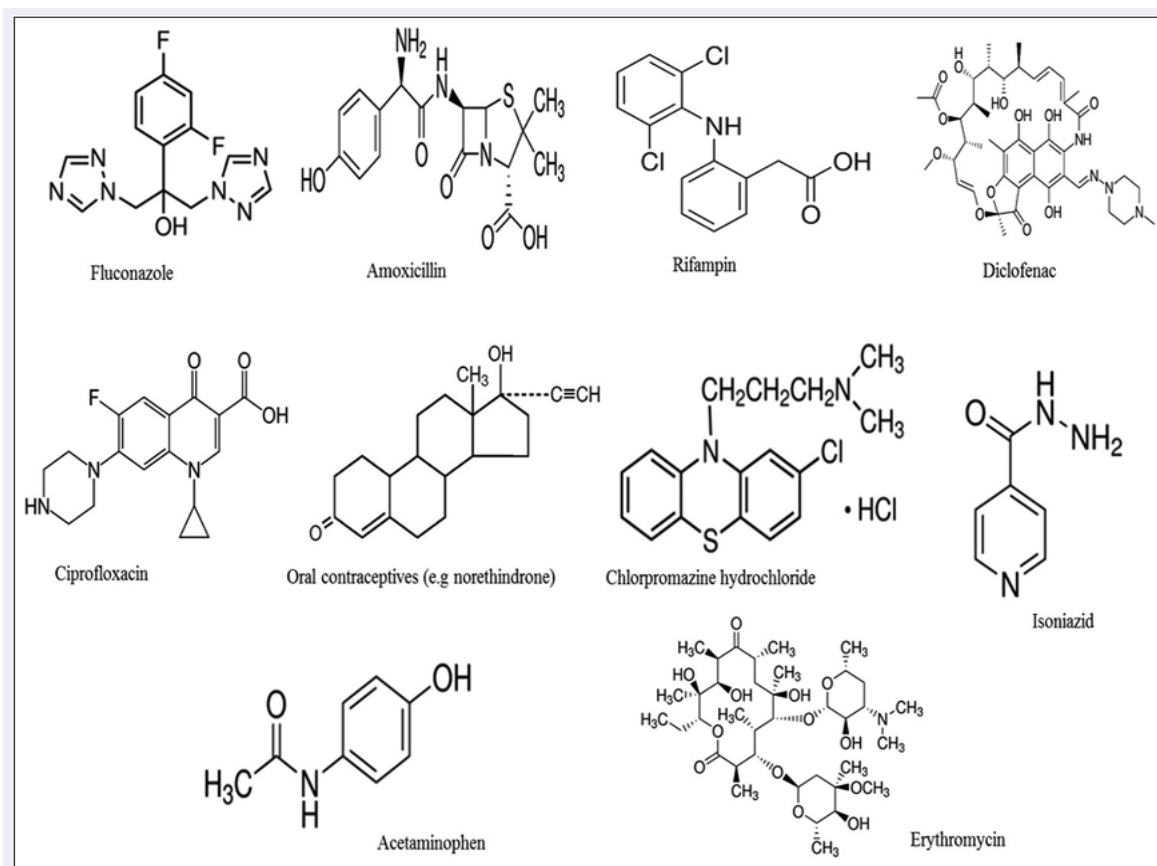
Drugs	Implication
Fluconazole	It leads to hepatitis; it increases the transaminase level, fulminant hepatic failure, and cholestasis
Amoxicillin	It moderates or brings about an increase in SGPT and SGOT levels, hepatic failure such as jaundice, acute cytolytic hepatitis, and hepatic cholestasis
Diclofenac	It elevates AST and ALT levels, jaundice, fulminant hepatitis, and liver necrosis
Rifampin	It leads to hepatitis, hyperbilirubinemia, and cholestasis Ciprofloxacin Elevation of SGOT alkaline phosphatase and SGPT levels occurs from cholestatic jaundice
Oral	Benign neoplasm, hepatic vein occlusion, and jaundice, contraceptives but rarely neoplasm of the liver
Chlorpromazine	It leads to infectious hepatitis with obstructive jaundice as a biomarker
Isoniazid	It elevates the serum transaminase level, severe and fatal hepatitis
Acetaminophen	It makes the cytochrome P-450-2E1 produce a toxic metabolite NAPQI that causes hepatic necrosis Erythromycin. It increases SGPT and SGOT concentration, and it also brings about hepatocellular hepatitis that are sometimes associated with it.

SGOT=Serum glutamic oxaloacetic  
transaminase, ALT=Serum alanine  
aminotransferase, AST=Aspartate  
aminotransferase, SGPT=Serum glutamic  
pyruvic  
NAPQI=N-acetyl-P-benzoquinoneimine

### Diseases of the Liver and Alcohol

In today's world, a major contributor to liver issues is drinking too much alcohol.[14] The liver is involved in alcohol metabolism, which impacts lipoprotein and lipid metabolism, which in turn links ethanol use to alcoholic liver disease. Additionally, cytochrome P450E1 is activated during the alcohol dehydrogenase-to-acetate conversion of ethanol to ROS. The numbers [29,30] The liver goes through a series of events that culminate in oxidative stress [Figure 2], which in turn causes damage

to the liver and alters the structural stiffness of the cell membrane, allowing cytosolic enzymes to seep into the circulation. As a result, elevated levels of cytosolic enzymes in the blood are the most prevalent biochemical indicator of liver injury.[31] In Both the cytoplasm and mitochondrial concentrations of aspartate transaminase (AST) and alanine transaminase (ALT) rise in injured liver cells. A change in the structure of the liver cell membrane, brought about by membrane leakage, leads to an elevation in serum hepatospecific enzymes. Furthermore, elevated blood bilirubin levels indicate a quickening of the erythrocyte degradation rate. Therefore, maintaining a healthy liver is critical to human wellbeing.[32] The process of alcohol-induced liver damage is shown in



### MEDICINAL PLANTS AS AN ALTERNATIVE TREATMENT

Metabolic diseases, including liver impairment, account for the vast majority of deaths and illnesses worldwide. The harmful effects of numerous allopathic medications on the liver have recently brought liver injury therapies into the spotlight on a worldwide scale. Therefore, researchers have paid a lot of attention to folkloric herbs that have hepatoprotective

properties for treating liver damage or illness. This is mostly because these herbs are therapeutic and have minimal toxicity. Animal studies have recently looked at the hepatoprotective effects of many traditional medicines. The positive effects of herbal medicine on human health have long been recognised and used in traditional medicine. Scientists have isolated several molecule types and investigated their pharmacological and

physicochemical characteristics. It is important to synthesise compounds and extracts correctly so that they may achieve their physiological aim and exert their pharmacological effects. The absorption and delivery of bioactive compounds may be impacted by factors including poor solubility and permeability.[32] However, to ensure their stability during the duration of usage, it is

important to assess the shelf life of herbal medications. Environmental factors such as heat, moisture, acidity, oxygen, and light speed up the degradation process. Herbal remedies include a wide variety of chemical components, including primary and secondary metabolites, carbohydrates, proteins, and lipids.[33]

Numerous efforts have been undertaken to discover new sources of hepatoprotective agents due to the adverse effects associated with synthetic medicines. on page 34 These days, the majority of hepatoprotective medications used to treat various liver illnesses are derived from plants, either in the form of individual plants or complex mixes of herbs. Rural residents, particularly in underdeveloped nations without appropriate modern healthcare facilities, rely heavily on folkloric herbs to enhance their quality of life.[1] The medicinal value of plants exceeds 70,000 species. While plants have gained popularity as natural treatment options, the science behind their preparation and dosage is often unclear. Despite their effectiveness and cost-effectiveness, it is imperative to prioritise plants with low toxicity. There are many herbal drugs on the market, but only a few bioactive ingredients have been proven to have antiviral, antioxidant, anticarcinogenic, antifibrotic, and anti-inflammatory effects.35 and 36

#### **IN VITRO AND IN VIVO HEPATOTOXICITY ASSAYS**

The choice of appropriate treatment for the liver disease relies solely on the suitability of the model the system preferred for hepatic damage. Although a number of prototypes exist to assess the hepatoprotective prospect of any chemical or plant extract, most of these models have limitations. Hence, it will be appropriate to combine these models for better results.[37] In a study carried out by Cerný *et al.*[38] *In vitro* assays such as hepatocyte cultures, perfused

hepatocytes study with pathophysiological damage caused by various chemical substance (e.g., hypoxia hepatotoxins, or anoxia assays in perfused immobilized hepatocytes were documented. Some of the reported *in vivo* models are presented in Table 2. The majority of these studies on the hepatoprotective role of some medicinal plants are still based on the laboratory experiments.

Most organisms have their own antioxidant-based defense mechanisms which combat the activity of free radical species. The protective role of the endogenous antioxidant system in humans is not always adequate when the free radical species are much greater than the available antioxidant, and hence, additional antioxidants from different sources become important. Various antioxidant agents of plant origin appear to be effective in scavenging free radicals that lead to liver injuries.[39] Phytochemicals such as phenolics, thiols, and carotenoids present in herbal plants protect the human body against oxidative damage by ROS.[17] Therefore, attention is being diverted to promising medicinal plants that have the hepatoprotective potential to treat different kinds of liver disease. The folkloric herbs for treating all kinds of diseases have been in existence since ancient times due to their therapeutic efficacy and safety, and several herbs have been investigated for their hepatoprotective potential for the treatment of different types of liver disorders.[14] Numerous herbal formulations have proved to be effective therapeutic agents against various kinds of liver disorders [Table 2], and this review mainly focused on available literature on those that have been confirmed around the globe to have hepatoprotective properties.

#### **SOME MEDICINAL PLANTS WITH HEPATOPROTECTIVE ACTIVITY**

##### *Dodonaea viscosa* (Sapindaceae)

*The flowering plant Dodonaea viscosa is a member of the soapberry family. Sapindaceae may be found all throughout the Southern Hemisphere, from the subtropics to the warm and tropical temperate zones of Africa, the Americas, and Australia.[47] Traditional healers in this area have used this plant, which*

they name "Sanatha," for decades to control their patients' diabetes.[132] The antihyperlipidemic and hepatoprotective effects of an aqueous: methanolic (70:30) *D. viscosa* leaf extract were recorded in a study by Ahmed et al., [47]. The rabbits were induced with diabetes by alloxan. The results showed that as compared to the control group, the experimental group had lower blood levels of TAG, total cholesterol, LDL-C, HDL-C, ALT, and AST. In addition, levels of HDL-C, AST, and ALT were significantly raised by the extract. These results demonstrate that *D. viscosa* leaf extract has hepatoprotective properties.

### **Phyllanthus muellarianus (Euphorbiaceae)**

The straggling, monoecious, glabrous, climbing shrub or small tree known as *Phyllanthus muellarianus* is widely distributed in many African countries, including Senegal, Uganda, Mali, Congo, Togo, South Africa, and Ivory Coast.[40] A wide range of medical conditions, including paralysis, fever, and bacterial infections, have been treated using extracts from this plant. On pages 133 and 134, A phytochemical analysis of an extract from *P. muellarianus* leaf showed the presence of phytochemical components that may be responsible for the therapeutic action, including furosin, isoquercetin, phaselic acid, corilagin, nitidine, geraniin, and gallic acid.13, 35, 136 The hepatoprotective ability of an aqueous extract of *P. muellarianus* leaf was studied in 2017 by Ajiboye et al. in relation to hepatocellular indices, proinflammatory factors, oxidative stress, and lipid peroxidation in Swiss albino mice that had been injured in the liver by *b*-acetaminophen.[40] The results demonstrated that the acetaminophen-induced changes in the ALT, ALP, AST, ALB, and TB were significantly reduced by the aqueous leaf extract ( $P < 0.05$ ), according to this research. The potential antioxidant properties of the water-based leaf extract may be due to its capacity to counteract the increase in these liver enzymes caused by acetaminophen, suggesting a protective benefit against

acetaminophen-induced liver damage. It has been shown that gallic acid, a phytochemical component of this plant extract and a well-known antioxidant, may reverse AST, ALT, and ALP, as well as acetaminophen-induced liver damage.[137]

Similarly, the rat liver showed a substantial reduction in the activities of SOD, GSH, CAT, G6PH, and GSH-Px when acetaminophen was used. Aqueous leaf extract from the plant under study considerably reduced the increase in levels of several compounds, including malondialdehyde, lipid hydroperoxides, fragmented DNA, protein carbonyl, and tumour necrosis factor- $\alpha$ . Because of its preventative properties, the plant was also determined to have promising future use as a dietary supplement.[40]

### **Aquilaria agallocha (Thymelaeaceae)**

This massive tree may reach heights of 60–80 feet and has a trunk that is three to four feet in diameter. Its original habitat is in South-east Asia. Similar to *Betula*'s usage of tree bark for writing, the bark is papery thin and was sometimes used for that purpose. The thin, leathery leaves may grow to be three inches in length. White blossoms accompany smooth, slender fruit that is one to two inches in length. *Aquilaria agallocha* is a plant with a wide range of pharmacological effects, including but not limited to: protecting against cancer, inflammation, diabetes, anxiety, ulcers, and seizures; alleviating pain; reducing fever; and antidiabetic, antihistaminic, antipyretic, laxative, antidiarrheal, antidiabetic, antihistaminic, sedative, antibacterial, and antimicrobial properties.[138] A study conducted by Alam et al. [72] shown that a 400 mg/ml ethanolic extract of *A. agallocha* (AAE) leaves protected the livers of Sprague-Dawley (SD) rats against PCM-induced hepatotoxicity. The findings demonstrated that AAE leaves have a hepatoprotective effect, as they prevented PCM-induced histopathological changes in the liver, increased ALB and total protein concentration, and significantly reduced AST, ALP, ALT, LDH, CHL, and TB in SD rats.[72]

**Table 2: Medicinal plants with hepatoprotective potentials**

Family	Name of the plant	Plant parts used	Extract used	Oral dose (mg/kg)	Hepatotoxicity inducing agents	Biochemical and histopathological parameter studied	Reference
Euphorbiaceae	<i>Phyllanthus muellarianus</i>	Leaves	Aqueous	400 mg/kg	Acetaminophen	ALP, ALT, AST, ALB, TB, CAT, SOD, GSH-Px, GSH SGOT, SGPT, ALP, CHL, TB, and TP AST, ALP, GGT, ALT, TBARS, and liver protein	[40]
Scrophulariaceae	<i>Picrorhiza kurroa</i>	Roots	Ethanol Alcohol	2.60 ml/kg	CCl <sub>4</sub> CCl <sub>4</sub>	ALT, ALP, and AST	[41]
Fabaceae	<i>Bauhinia variegata</i>	Stem barks	Alcohol Methanol	100 and 200 mg/kg	CCl <sub>4</sub> CCl <sub>4</sub> CCl <sub>4</sub> PCM Alloxan	ALT, TBARS, and liver protein	[42]
Rubiaceae	<i>Galium aparine</i>	Whole plant	Methanol/ethyl acetate	2 ml/kg	CCl <sub>4</sub> PCM CCl <sub>4</sub>	ALT, AST, and ALP	[43]
Cannaceae	<i>Canna indica</i>	Ficus	Ethanol Methanol		Country-made liquor		
Moraceae	<i>Ficus cordata</i>	Aerial parts					
Zingiberaceae	<i>Curcuma longa</i>	Roots	Methanol	100 and 200 mg/kg	PCM	SGPT, SGOT, TB, CAT, GSH, LPO LDH	[44]
Sapindaceae	<i>Dodonaea viscosa</i>	Roots	Methanol	400 mg/kg			[45]
Asteraceae	<i>Eclipta prostrata</i>	Rhizome	Ethanol and aqueous	600 mg/kg		ALT, ALP, and AST	[46]
Cyatheaceae	<i>Cyathea gigantea</i>	Leaves	Ethanol	500 mg/kg		LDLC, ALT, HDL STG, and TC	[47]
Araceae	<i>Alocasia macrorrhizos</i>	Fresh leaves	Methanol, petroleum ether, chloroform, and acetone, and aqueous	100 and 200 mg/kg	CCl <sub>4</sub> TAA d-GalN/LPS CCl <sub>4</sub> CCl <sub>4</sub>	ALT, AST, and serum bilirubin SGPT, SGOT, ALP, TB, TP	[48]
Nyctaginaceae	<i>Boerhavia diffusa</i>	Leave and tuber				Serum ALT and AST	[49]
Apocynaceae	<i>Leptadenia pyrotechnica</i>	Roots	Methanol	200 mg/kg		SGPT, SAP, TGs, and total lipid levels	[17]
Asclepiadoideae		Whole plant				SGPT, TB, ALP, and SGOT	[50]
Areaceae			Methanol Ethanol	200 and 400 mg/kg			
Asteraceae				150 ml/kg			
Cactaceae	<i>Tylophora</i>	Leaves Fruit	Aqueous		CCl <sub>4</sub> CPF Ethanol CCl <sub>4</sub>		[7]
Rutaceae	<i>Phoenix dactylifera</i>	Aerial parts	Methanol		CCl <sub>4</sub> /ethanol		
	<i>Tridax procumbens</i>					SGPT, ALP, SGOT, and bilirubin content	
	<i>Opuntia ficus-indica</i>	Leaves		200 and 300 mg/kg	PCM	TBAST, ALT, and ALP	[51]
	<i>Clausena lansium</i>	Stem bark		300 mg/kg	INH and RIF	AST, LDH, ALT, ALP, GGT, TB, and TBARS	[52]
Apiaceae			Methanol	300 mg/kg		AST, ALT, creatinine, urea, and uric acid	[53]
Cactaceae			Aqueous			Reduction in phenobarbitone, sleeping time and serum liver protein, serum AST, ALT, and ATP.	[54]
Rosaceae	<i>Apium graveolens</i>	Seeds Stem	Alcohol N/A	2, mL/kg			[55]
Vitaceae	<i>Opuntia ficus-indica</i>	Aerial part	Ethanol Alcohol	100 and 200 mg/Kg		SGOT, SGPT, SALP, TP, TA, and GSH ALAT, ASAT, ALP, LDH, CHL, and albumin	[56]
Pandanaceae	<i>Agrimonia eupatoria</i>	Leaves Aerial part				AST and ALT	[57]
Rhamnaceae	<i>Vitis vinifera</i>	Roots					
	<i>Rheum palmatum</i>	Roots		250 mg/Kg			
	<i>Pandanus odorifer</i>			1500 mg/kg			
	<i>Ziziphus oenoplia</i>					AST and ALT N/A	[58]
				100 and 300 mg/kg		SGOT, SALP, SGPT, TB, and TGA SGOT, SGPT, SALP, SB, SOD, CAT, GST, and GPx	[59]
				125 mg/kg			[60]
				25 and 100 mg/kg			[61]
				200 and 400 mg/kg			[62]
				150 and 300 mg/kg			

Family	Name of the plant	Plant parts used	Extract used	Oral dose (mg/kg)	Hepatotoxicity inducing agents	Biochemical and histopathological parameter studied	Reference
Asteraceae	<i>Cichorium intybus</i>	Leaves	Ethanol	50, and	CCl <sub>4</sub>	ALT, AST, and ALP	[63]
Betulaceae	<i>Corylus avellana</i>	Leaves Bark	Aqueous	100 mg/kg	CCl <sub>4</sub> and	GPT and GOT	[64]
Lauraceae	<i>Cinnamomum cassia</i>	Seeds Gums	Ethanol	NA	acetaminophen		
Apiaceae	<i>Anethum graveolens</i>	Edible portion (seed coats and juice)	NA	40 mg/kg	Dimethylnitrosamine	TP, albumin, TB, direct bilirubin, GOT, GPT, and ALP	[65]
Anacardiaceae	<i>Pistacia lentiscus</i>	Flower	Acetone	500 and 1000 l/kg	CCl <sub>4</sub> CCl <sub>4</sub>	SGPT, SGOT, and ALP	[66]
Rosaceae	<i>Punica granatum</i>	Aerial parts	Aqueous	400mg/kg	INH and RIF	AST, ALT and MDA, GSH, GPx, GST, GR, SOD, and CAT	[67]
Cucurbitaceae	<i>Rosa damascena</i>	Fruits	Methanol		Acetaminophen	AST, ALT, and LDH	[68]
Muntingiaceae	<i>mill</i>	Leaves	Methanol	250, 500 and 1000 mg/kg	CCl <sub>4</sub>	AST, ALT, ALP, LDH, ALBTB, urea and creatinine, TBARS, and GSH	[69]
Thymelaeaceae	<i>Cucurbita maxima</i>	Root	NA		Acetaminophen PCM	SGPT, SGOT, ALP, TP, and TB	[70]
Berberidaceae	<i>Muntingia calabura</i>	Whole plant	Hydroalcohol	250 and 500 mg/kg	CCl <sub>4</sub> CCl <sub>4</sub>	AST, ALT, and ALP	[71]
Apiaceae	<i>Aquilaria malaccensis</i>	Roots	Methanol	50, 250, and 500 mg/kg	Acetaminophen/CCl <sub>4</sub>		
Asteraceae	<i>Coptidis rhizoma</i>	Edible root and shoot	Hydroalcoholic acid	400 mg/kg	Ethanol		
Asteraceae	<i>Cynara scolymus</i> L.	Root	Methanol	120 mg/kg	CCl <sub>4</sub>	AST, ALT, LDH, ALP, bilirubin, CHL, TP, and ALB	[72]
Euphorbiaceae	<i>Calendula officinalis</i>	Root	Methanol	900 mg/kg	TAA	ALT, AST, and SOD	[73]
Fabaceae	<i>Taraxacum officinale</i>	Aerial parts	Ethanol	500 mg/kg	TAA	ALT, ALP, AST, GSH, and CAT	[74]
Euphorbiaceae	<i>Tragopogon porrifolius</i>	leaves	Methanol	250 mg/kg	CCl <sub>4</sub>	AST, and LDH	[75]
Fabaceae	<i>Baliospermum montanum</i>	Leaves	Ethanol	2000 mg/kg	H <sub>2</sub> O <sub>2</sub> ; CCl <sub>4</sub> CCl <sub>4</sub>	TBARS, GST, GSH, SOD, CAT, GR, and GPx	[76]
Rutaceae	<i>Tephrosia purpurea</i>	Leaves, bark	Methanol	500 mg/kg	CCl <sub>4</sub> CCl <sub>4</sub>	CAT, SOD and GSTAST, ALT, and LDH	[77]
Acanthaceae	<i>Alchornea cordifolia</i>	Leaves	Methanol	300 mg/kg	PCM	GOT, GPT, ALP, TB, TC, TB, and albumin	[78]
Asteraceae	<i>Trigonella foenum-graecum</i> L.	Leaves	Ethanol	100 mg/kg		AST, GSH, ALT, ALP, TB, GGT, and MDA	[79]
Fabaceae	<i>Glycosmis pentaphylla</i> Corr.	Seeds	Methanol	100 mg/kg		SGOT/AST, SGPT/ALT, ALP, and TB	[80]
	<i>Andrographis lineata</i> Nees			500 mg/kg		ALT, AST, ALP, and GGT	[81]
	<i>Wedelia chinensis</i> L.			845 mg/kg		ALT/SGPT, AST/SGOT, CHL, bilirubin, and glucose	[82]
	<i>Cassia fistula</i>			200 mg/kg		SGOT, SGPT, and ALP	[83]
				200 and 400 mg/kg		AST, ALT, ALP, protein, and bilirubin	[84]
						SGOT, SGPT, ALP, and bilirubin	[85]



<i>Fabaceae</i>	<i>Bauhinia racemosa</i>	Bark	Stem	Methanol	NA	CCl4 and PCM	CCl4	SGPT, SGOT, SOD, GSH, and TBARS	AST, ALT, and GGT	[86]
<i>Fabaceae</i>	<i>Lam. Bauhinia</i>	bark		Methanol		PCM		ALT, ALP, and GGT		[87]
<i>Scrophulariaceae</i>	<i>variegata L.</i>	Aerial parts		Ethanol	100 and	Acetaminophen	CCl4	ALT, GGT, AST, and ALP		[88]
<i>e Phyllanthaceae</i>	<i>Scrophularia</i>		Whole plant	Methanol	200 mg/kg	Ethanol		Cytochrome P450		[89]
<i>Phyllanthaceae</i>	<i>hypericifolia</i>	Fruits		NA	250 and			CYP2E1 protein GSH		[89]
<i>Liliaceae</i>	<i>Phyllanthus urinaria</i>	Fresh bulbs		Aqueous	500 mg/kg	Rifampicin		ALT, ALP, AST, and TB NA		[90]
<i>Moraceae</i>	<i>Allium cepa</i>				200 mg/kg					[90]
		Leaves, fruit, and roots		Petroleum ether extract, aqueous extract, and methanolic extract	100, 300 and 600 mg/kg	Dimethoate				[91]
<i>Rhamnaceae</i>	<i>Ficus carica</i>	Leaves		Methanol	NA			SGOT, TBARS, SGPT, GSH, SOD, tocopherol, HDL, LDL, CHOL, TL, TGA		[92]
								Induce apoptosis of hepatic stellate cells (HSCs)	AST, ALT, ALP	[93]
<i>Lamiaceae</i>	<i>Ziziphus mucronata</i>			Ethanol	200 mg/kg	Mixture of cholesterol and cholic acid with coconut oil	CCl4			[93]
<i>Mahvaceae</i>	<i>Salvia miltiorrhiza</i>	Dried pulverized roots	Flower	Aqueous		CCl4				[94]
	<i>Hibiscus rosasinensis</i>	Leaves		Aqueous and ethanol	50 mg/kg	CCl4		AST, TP ALP, and ALT, TB		[94]
<i>Loranthaceae</i>	<i>Dendrophthoe falcata</i>	Dried aerial parts	Fruiting bodies and mycelia	Aqueous extract and ethanol	80, 160 and 240 mg/kg	CCl4	CCl4	induced elevation of expression of hepatic mRNAs, i.e., MMP-9, TNF- $\alpha$ , KLF-6, and TGF- $\beta$ 1 levels		[15]
<i>Asteraceae</i>	<i>Bidens pilosa</i>					CCl4	PCM STZ			[95]
<i>Fomitopsis Deae</i>	<i>Antrodia cinnamomea</i>					CCl4	CCl4			[95]
<i>Cyperaceae</i>		Leaves		Methanol	15 mg/kg	CCl4		SGOT, SGPT, ALP		[96]
<i>Malvaceae</i>		Aqueous extract	Dried root	Aqueous and methanol	1250 mg/kg	PCM	PCM	ALT, AST, and ALP		[97]
<i>Polygonaceae</i>	<i>Cyperus rotundus</i>							ALT, AST, HA, and laminin (LN)	sGPT, serum glutamic ALP, TBARS, and GSH	[98]
<i>Araceae</i>	<i>Hibiscus sabdariffa</i>	Tubers	Leaves	Ethanol	200 mg/kg			ALP and GGT		[99]
<i>Apiaceae</i>	<i>Rheum palmatum</i>	Leaves	Dried seeds	Ethanol	NA					[99]
<i>Fabaceae</i>	<i>Amorphophallus paeonifolius</i>	Whole plants		Methanol						[100]
<i>Fabaceae</i>	<i>Petroselinum crispum</i>	Leaves		Aqueous methanol	400 mg/kg			Induce apoptosis of hepatic stellate cells (HSCs)	SGOT, SGPT, and ALP	[100]
<i>Oxalidaceae</i>	<i>Loranthus parasiticus</i>	Aerial parts			300 mg/kg			TBARS, SOD, CAT, and GSH		[101]
<i>Fabaceae</i>	<i>Trigonella foenum-graecum</i>				200 mg/kg			TB, DB, ALP, and AST		[102]
<i>Malvaceae</i>	<i>Tephrosia purpurea</i>				100 mg/kg					[103]
	<i>Oxalis corniculata</i>				20-100 mg/kg					[103]
	<i>Indigofera tinctoria</i>				50-200 mg/kg					[104]
	<i>Alcea rosea</i>				100 mg/kg, 75, 150, 300 mg/kg, 200 mg/kg					[94]
										[105]
										[106]
										[107]

<i>Fabaceae</i>	<i>Cajanus cajan</i>	Whole plant	Methanol	NA	CC14	SGOT, CHL, and SGPT	[108]
<i>Solanaceae</i>	<i>Cestrum nocturnum</i>	Leaves	Aqueous ethanol	250 and	PCM PCM CCI4 CCI4	SGOT, SGPT, ALP,	[109]
<i>Convolvulaceae</i>	<i>Convolvulus</i>	Whole plant	Ethanol	500 mg/kg	Galactosamine/ lipopolysaccharide	AST, ALT, and LDH	[110]
<i>Fabaceae</i>	<i>arvensis Glycyrrhiza</i>	Roots		200 and	CCI4	ALP, ALP, AST, and TB	[110]
<i>Convolvulaceae</i>	<i>glabra</i>		Aqueous	500 mg/kg	PCM PCM	SOD, GST, CAT, GSH, and GSH-Px	[111]
<i>Malvaceae</i>	<i>Ipomoea</i>	Leaves	Hydroalcohol	2 mg/kg		ALP, SGOT, AST, CHL, ALT, SGPT	[28]
<i>Ramunculaceae</i>	<i>staphylina Malva</i>		Methanol		PCM and CCI4 CCI4	ALT, TP, AST, and ALP	[112]
<i>Oleaceae</i>	<i>parviflora</i>	Whole plant	Alcohol Ethanol	200 mg/kg	CCI4 CCI4	ALP, ALT, TB, AST, and TP	[113]
<i>Polygonaceae</i>		Seeds	Aqueous/ methanol	250 and	CCI4 PCM PCM CCI4	GOT, GPT, CAT, SOD, and GPx	[114]
<i>Amaranthaceae</i>	<i>Nigella sativa</i>	Whole plant	Aqueous/ methanol	500 mg/kg		ALP, ALT, TB, AST, and TP	[113]
<i>Lamiaceae</i>	<i>Fraxinus</i>	Whole plant	Aqueous/ methanol	NA	PCM	GOT, GPT, CAT, SOD, and GPx	[115]
<i>Boraginaceae</i>	<i>rhynchophylla</i>	Leaves	Aqueous/ether			ALP, ALT, TB, AST, and TP	[116]
	<i>Rumex dentatus</i>	Leaves	Ethanol	100 and		SGPT, ALP, ALT, SGOT, AST, TP, and TB	[117]
<i>Vitaceae</i>		Leaves		500 mg/kg		AST, CAT, TB, ALT, and TP	[118]
<i>Acanthaceae</i>	<i>Suaeda fruticosa</i>	Leaves	Ethanol Aqueous	250 and	PCM CCI4	SGPT, SGOT, ALP, and TB	[119]
		Leaves		500 mg/kg	CCI4	MDA, GSH, protein, bilirubin, SGOT, ALP, and SGPT	[120]
<i>Lamiaceae</i>		Roots	Methanol	500 and		ALT, and ALP	[121]
<i>Fabaceae</i>	<i>Thymus linearis</i>	Roots	Ethanol	750 mg/kg	PCM CCI4	ALT, ALP, and AST	[122]
<i>Plumbaginaceae</i>		Fresh leaves	Methanol		PCM	TB, SGPT, SGOT, and ALP	[123]
<i>Salicaceae</i>	<i>Trichodesma</i>	Leaves	Ethanol	250 and		albumin, TB, TG, urea, creatinine, TB, TBARS, and GSH	[29]
	<i>sedgwickianum</i>	Aerial parts		500 mg/kg		AST, SGOT, SOD, GSH, ALP, SOD, and TP	[117]
	<i>Vitis vinifera</i>	Flowers		400 mg/kg	PCM and azithromycin CCI4	AST, CAT, TB, ALT, and TP	[118]
<i>Bignoniaceae</i>			Aqueous/ ethanol			SGPT, SGOT, ALP, and TB	[119]
	<i>Hygrophila auriculata</i>	Aerial parts		200 mg/kg		ALP, ALT, and AST	[124]
<i>Anacardiaceae</i>	<i>Ocimum gratissimum</i>		Ethyl acetate			ALP, ALT, and AST	[125]
<i>Scrophulariaceae</i>	<i>Bauhinia purpurea</i>	Bark	Ethanol	100 mg/kg		ALP, ALT, and AST	[126]
<i>e</i>	<i>Plumbago zeylanica</i>	Whole plant		40 mg/kg		ALP, ALT, and AST	[127]
<i>Verbenaceae</i>	<i>Salix caprea</i>	Leaves	Alcohol	50 and		ALP, ALT, and AST	[128]
		Fruits	Methanol	250 mg/kg		ALP, ALT, and AST	[129]
<i>Lamiaceae</i>		Stem barks		300 mg/kg		ALP, ALT, and AST	[130]
<i>Mimosaceae</i>	<i>Tecomella undulata</i>		Aqueous Ethanol			ALP, ALT, and AST	[13]
			Ethyl alcohol	150 mg/kg		ALP, ALT, and AST	[13]
<i>Rubiaceae</i>	<i>Pistacia integerrima</i>					ALP, ALT, and AST	[108]
<i>Cupressaceae</i>	<i>Scoparia dulcis</i>					ALP, ALT, and AST	[108]
<i>Oleaceae</i>						ALP, ALT, and AST	[108]
	<i>Stachytarpheta jamaicensis</i>			100 and		ALP, ALT, and AST	[108]
	<i>Ocimum tenuiflorum</i>			200 mg/kg		ALP, ALT, and AST	[108]
	<i>Mimosa pudica</i>			NA		ALP, ALT, and AST	[108]
	<i>Kohautia grandiflora</i>			500 and		ALP, ALT, and AST	[108]
	<i>Juniperus communis</i>			1000 mg/kg		ALP, ALT, and AST	[108]
	<i>Fraxinus rhynchophylla</i>			200 mg/kg		ALP, ALT, and AST	[108]
				200 mg/kg		ALP, ALT, and AST	[108]
				200 mg/kg		ALP, ALT, and AST	[108]
				300 mg/kg		ALP, ALT, and AST	[108]
				200 mg/kg		ALP, ALT, and AST	[108]
				400 mg/kg		ALP, ALT, and AST	[108]
<i>Family</i>	<i>Name of the plant</i>	<i>Plant parts used</i>	<i>Extract used</i>	<i>Oral dose (mg/kg)</i>	<i>Hepatotoxicity inducing agents</i>	<i>Biochemical and histopathological parameter studied</i>	<i>Reference</i>
<i>Saururaceae</i>	<i>Saururus chinensis</i>	Whole plant	Ethanol	70 mg/kg	CCI4	AST, ALT, ALP, CHL, SOD, CAT, MDA, and GSH	[131]

SGOT=Serum glutamic oxaloacetic transaminase, SB=Serum bilirubin, SOD=Superoxide dismutase, GST=Glutathione S-transferase, ALP=Alkaline phosphatase, GPT=Glutamic pyruvic transaminase, ALT=Serum alanine aminotransferase, AST=Aspartate aminotransferase, SALP=Serum alkaline phosphatase,

MDA=Malondialdehyde content, GSH=Glutathione, GSH-Px=Glutathione peroxidase, TG=Triglycerides, GPT=Glutamic pyruvic transaminase, TB=Total bilirubin, TBARS=Lipid peroxidation (thiobarbituric acid reactive substance), GSH=Reduced glutathione, ALB=Albumin, GR=Glutathione reductase, SALP=Serum alkaline phosphatase, GGT=Gamma glutamyl transferase, SALT=Serum aspartate amino transaminase, GPx=Glutathione peroxidase,

GR=Glutathione reductase, CHL=Cholesterol, LDH=Lactate dehydrogenase, SGPT=Serum glutamate pyruvate transaminase, TP=Total protein, GOT=Glutamic oxaloacetic transaminase, CAT – catalase, TB=Total bilirubin, NA=Not applicable, CPF=Organophosphorous insecticide chlorpyrifos, TAA=Thioacetamide, d-GalN/LPS=d-galactosamine/lipopolysaccharide, INH=Isoniazid, RIF=Rifampicin, N/A=Not available

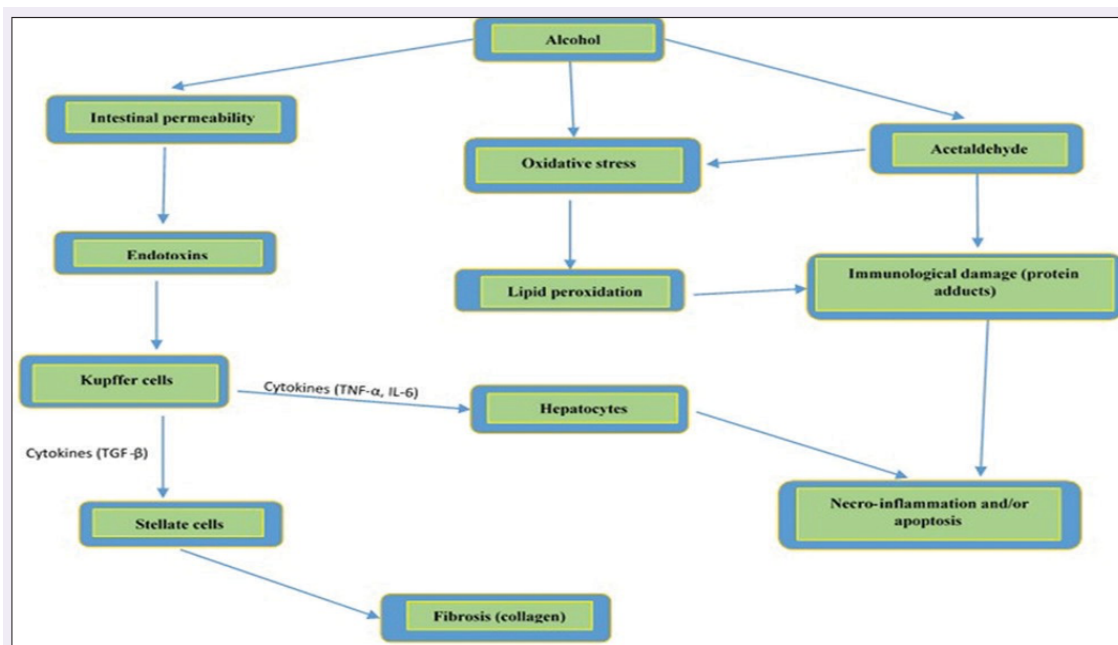


Figure 2: Mechanism of alcohol-induced hepatotoxicity[2]

### Salix caprea (Salicaceae)

One of *Salix caprea*'s family names is Salicaceae.[29] One of the most common types of willow in Europe, Western and Central Asia, this tree is also known as great willow, pussy willow, or goat willow. Past research has shown that the plant has several biological potentials, including antioxidant, anti-inflammatory, and anti-inflammatories properties. at the number 47,139 Traditional healers have used *S. caprea* to cure a wide range of illnesses in both humans and animals. It is used most often to alleviate symptoms such as fever, headaches, stomachaches, and constipation.[29] The plant extract was shown to contain bioactive components, namely flavonoids and phenolic acids, according to qualitative analysis. Salignin and catechins were among the phenolic chemicals found,

while quercetin, rutin, and luteolin-7-glucoside were flavonoids. the number 140

In CCl4-induced liver damage, Wahid et al. evaluated the hepatoprotective efficacy of an ethanolic extract of *Salix subserrata* Willd flower.[29] The ethanolic extract of this plant significantly reduced serum enzyme levels, suggesting that it may repair CCl4-induced damage to liver tissue or restore plasma membrane integrity. This plant extract may have shown antioxidant activity because of its hepatoprotective function, which prevented CCl4 free radical metabolites from damaging liver cells by preventing lipid peroxidation, membrane destabilisation, and subsequent cell death.

Caesalpinia crista (Fabaceae)

A wide shrubby perennial climber widespread all throughout India in the plains, deserts, coastal regions, and hills up to a height of 1000 m, *Caesalpinia crista* is a member of the Fabaceae family, a genus of flowering plants in the legume family. It is often known as *Karanja* in Hindi.[141] The bioactive substances found in *C. crista* include glycosides, alkaloids, saponins, and flavonoids. The medical properties of this plant include actions against inflammation, malaria, jaundice, worms, diabetes, periodontal disease, and fever. In references [61,142], Mishra et al. evaluated the hepatoprotective efficacy of ethanolic extracts of *C. crista* leaves in rats subjected to PCM-induced hepatotoxicity. The positive control group showed no change in levels of TB and serum marker enzymes or TGA when treated with ethanolic extract at doses of 200 or 400 mg/kg. In contrast, the treated group showed a significant drop in these levels. The ethanolic extracts of *C. crista* leaves showed promising hepatoprotective effects against PCM-induced liver damage in rats, according to these results.[61]

#### *Alocasia indica* (Araceae)

Southern India, West Bengal, Assam, and Maharashtra are among the tropical and subtropical zones where *Alocasia indica* is widely grown. This herbaceous perennial reaches a height of about 5 metres.[17] Because it is both inexpensive and widely accessible, the edible tuber portion of the *A. indica* plant is often eaten as a common vegetable. Traditional medicine traditionally employs *A. indica* for the treatment of conditions involving the spleen and the abdomen. Although their study is focused on the nonedible leaf section of the plant, the edible tuber component is extensively used as a vegetable by the Indian people. The hepatoprotective effect of *A. indica* tuber extracts, both ethanolic and aqueous, was tested against CCl<sub>4</sub>-induced liver damage in male Albino Wistar rats by Pal et al. [14]. Their results showed that biochemical examination in both water and ethanolic extracts identified many pharmacological components including tannins, flavonoids, alkaloids, glycosides, and saponins. The antioxidant capacity and flavonoid and phenolic content were both found to be higher in the ethanolic extract compared

to the water-based one. In an *in vivo* experiment, both the water and alcohol extracts showed strong hepatoprotective effects. This study's findings suggest that this plant extract may be useful as an antioxidant in the development of medications for liver illnesses..

#### *Opuntia ficus-indica* (Cactaceae)

As a house-trained crop plant crucial to agricultural economies spread across the arid and semiarid regions of the globe, *Opuntia ficus-indica* is a species of cactus that is often employed for fruit production and belongs to the Cactaceae family. The most likely place of origin is Mexico. When herbaceous plants and water are scarce, it is common practice to utilise it as a vegetable fodder resource for cattle.[143] Instead than studying the fruit, most scientific medical research focuses on the leaves (cladodes). The ability of an aqueous extract (2 mL/kg) from cactus cladodes to protect the liver from CCl<sub>4</sub>-induced toxicity in male Wistar rats was investigated.[54] The results showed that compared to the group treated with CCl<sub>4</sub>, which caused hepatotoxicity, the group treated with aqueous extract of *O. ficus-indica* had significantly lower levels of AST and ALT..

#### *Cyathea gigantea* (Cyatheaceae)

The tree fern *Cyathea gigantea* (Wall. ex. Hook.) is native to damp, open habitats in Nepal, Western Java, Northeastern to Southern India, and Sri Lanka. The maximum height that this plant may reach is 20 metres. 144 in all The hepatoprotective efficacy of a methanolic extract of *C. gigantea* leaves on PCM-induced toxicity in Wistar Albino rats was shown by Kiran et al. [49]. Hepatic histological and biochemical damage was seen in the experimental rats after PCM intoxication. However, the liver damage was reversed by restoring the structural integrity of the plasma membrane, and the elevated levels of ALP, serum glutamic oxaloacetic transaminase, TB, and serum glutamate-pyruvate transaminase were decreased as a result of treatment with methanolic *C. gigantea* leaf extract. According to the results of the phytochemical screening, *C. gigantea* leaf extract contains phenols, triterpenes, saponins, sterols, and flavonoids. Its hepatoprotective activity may be due to these bioactive components. The rat PCM-induced

hepatotoxicity model was used to determine *C. gigantean's* hepatoprotective capabilities.

### Phoenix dactylifera (Arecaceae)

In the northern regions of Nigeria, Middle Eastern nations, and Arabian countries, the date palm, or *Phoenix dactylifera*, is frequently used to cure liver diseases and their symptoms, including jaundice.[52] The fruit extract shields the liver from harmful substances including alcohol, according to earlier research.[145] In a study on male rats, Okwuosa et al.[52] examined the possible hepatoprotective effects of date palm (*P. dactylifera*) fruit extracts in TAA-induced toxicity. Because the levels of hepatocellular enzymes were lower in the test groups compared to the TAA-induced group, the researchers concluded that the plant's methanolic fruit extract had strong hepatoprotective capabilities. The extract may have reversal potential for plasma membrane damage since it may counteract the increase in serum bilirubin and ALP caused by TAA.[52] Plant extracts were found to include tannins, flavonoids, saponins, terpenoids, carbs, steroids, proteins, and glycosides, according to qualitative screening. There have been reports that flavonoids may stabilise the cell membrane.[146] It is reasonable to assume that the flavonoid content of *P. dactylifera* extract is responsible for its membrane stabilising activity. Unfortunately, this research did not examine the biochemical mechanism by which *P. dactylifera* exerts its hepatoprotective effects. It was thought that the bioactive ingredient  $\beta$ -sitosterol in the fruit extract was the one responsible for its activity.[147] It was also thought that flavonoids' hepatoprotective effects were due to their ability to inhibit cytochrome P450 aromatase.[148] Likewise, Al-Qarawi et al. [149] confirmed that extracts from the flesh

and pits of dates (*P. dactylifera* L.) had a hepatoprotective effect against CCl<sub>4</sub>-induced toxicity in rats.

### Convolvulus arvensis (Convolvulaceae)

The creeping weed *Convolvulus arvensis* is native to Asia. The number 150 This kind of plant is a member of the family Convolvulaceae. When not climbing, the plant may spread out into thick carpets 5 cm thick; it is also often used as a laxative. Skin conditions, coughs, jaundice, and the flu may all benefit from the plant extract. Furthermore, it has the potential to alleviate inflammation, edoema, and aching joints. The hepatoprotective efficacy of an ethanolic extract of *C. arvensis* at doses of 200 and 500 mg/kg in rats subjected to PCM-induced toxicity was recently shown by Ali et al. [110]. The elevated levels of TB and hepatic enzymes were significantly reduced ( $P < 0.05$ ) in rats treated with PCM-induced ethanolic extract of *C. arvensis*. Quercetin and kaempferol were the primary phytochemical components of *C. arvensis*. One flavonoid with proven hepatoprotective properties is quercetin. The number 151

### Bioactive Molecules With Hepatoprotective Potentials

Past research has led to the isolation of several plant biomolecules with intriguing hepatoprotective properties. Humans have not yet been the subjects of comprehensive clinical studies with these pure substances. The presence of biomolecules such as resveratrol, curcumin, silymarin, glycyrrhizin, and quercetin gives some of the bioactive compounds their various biological properties, which include antiviral, antioxidant, anticancer, antiaging, antifibrotic, antidiabetic, and anti-inflammatory potentials (Table 3).

**Table 3: Examples of reported phytochemical compounds with hepatoprotective potential**

Phytochemical compounds	Plants
Glycyrrhizin	<i>Glycyrrhiza glabra</i>
Resveratrol	<i>Hygrophila auriculata</i>
Curcumin	<i>Curcuma spp.</i>
Colchicine	<i>Colchicum autumnale</i>
Silymarin (silybin)	<i>Silybum marianum</i>
Quercetin	<i>Hibiscus vitifolius</i>
Fumaric acid	<i>Sida cordifolia</i>

Coumarins	<i>Artemisia abrotanum</i>
Schizanthrin A	<i>Schisandra chinensis</i>
Kutkoside	<i>Picrorhiza kurroa</i>
Catechin	<i>Anacardium occidentale</i>
Papyriogenin	<i>Tetrapanax papyrifer</i>
Cronin	<i>Gardenia jasminoides</i>
Syringopicroside	<i>Syringa oblata</i>
Piceid	<i>Polygonum cuspidatum</i>
Gomishins	<i>Schisandra chinensis</i>
Saikosaponin	<i>Bupleurum falcatum</i>
Cosmosiin	<i>Cupressus sempervirens L.</i>
Patuletin	<i>Ficus ingens</i>

## CONCLUSIONS AND FUTURE PROSPECTS

Even though everyone is very concerned about their health, health problems have become a major issue in our society. Liver disorders and injuries are among the most prevalent medical problems worldwide, despite the liver's vital role in the body. Liver damage may be caused by a variety of things, the most common of which include poor dietary habits, excessive alcohol intake, herbal supplements, microbial infections, autoimmune illnesses, malignancies, metabolic diseases, and drug addiction. Therefore, it is crucial to safeguard the liver against the dangers listed above.

However, given that current therapy for treating various liver illnesses is either insufficient or linked with negative effects on renal function, the need for the discovery of new therapies that effectively cure liver damage has become urgent. Consequently, the development of novel hepatoprotective medicines derived from plants is imperative. Their antioxidant-related characteristics and hepatoprotective actions are the basis of most plant-based medications used to treat liver disorders. In order to treat liver illnesses with innovative medications that have minimal adverse effects on the kidney, these are the key scientific principles that are founded on. Hence, more research is required to assess the chances of developing more powerful hepatoprotective medications from novel candidate phytochemicals.

Due to the fact that half of the population in poor nations relies on herbal treatments to cure liver diseases, these therapies have become famous across the world. The majority of commercially available herbal extracts have

shown encouraging results in alleviating the signs and symptoms of liver damage or illness. Even yet, there has been no proof of the herbal extracts' scientific validity; consequently, further study is required, particularly in this field, to define protocols for the safe and effective manufacture and administration of herbal extracts. Also, it's crucial to put these herbal remedies through preclinical testing before putting them through clinical trials. The therapeutic efficacy of these all-natural herbal remedies may then be assessed, and the established dose regimen from clinical trials can inform future medication development and delivery. In addition, many important medications for treating various disorders may be made accessible using the conventional medical method to drug discovery and design. It takes a lot of effort and money to isolate active principles and turn them into medications. The availability of plant-based medications, namely those derived from individual or mixed plant extracts, should be prioritised in the treatment of liver illnesses, particularly those that aim to restore the hepatic cell membrane's structural integrity. In most cases, a single plant extract will not be able to cure every kind of liver illness. Because of this, it may be required to create a herbal combination using extracts from two or more plants in order to increase the treatment's effectiveness. Furthermore, further study, particularly toxicity tests, should be conducted to guarantee the plant combination is safe to use. This is because it is quite probable that one of the plant extracts may be poisonous, which would undermine the effectiveness of the other extracts in the mixture. Also, traditional healers

in developing nations need to be educated on proper hygiene practices when preparing plants for use, as the majority of plant extracts are used by impoverished rural residents. Contamination must be prevented or eliminated when herbal extracts are prepared.

In conclusion, more research into the structural alterations of the active principles produced from herbal extracts using computational chemistry methods is required in order to develop plant-based hepatoprotective medicines that are more successful.

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